

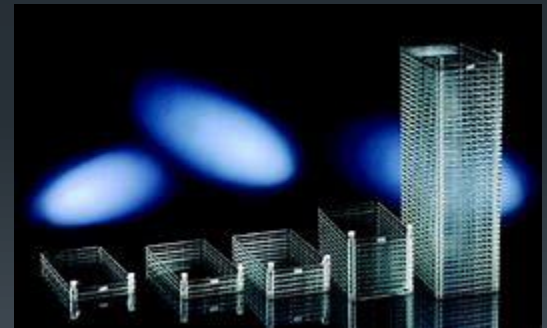
# AAV-Process Flow-Chart

Mason Snead



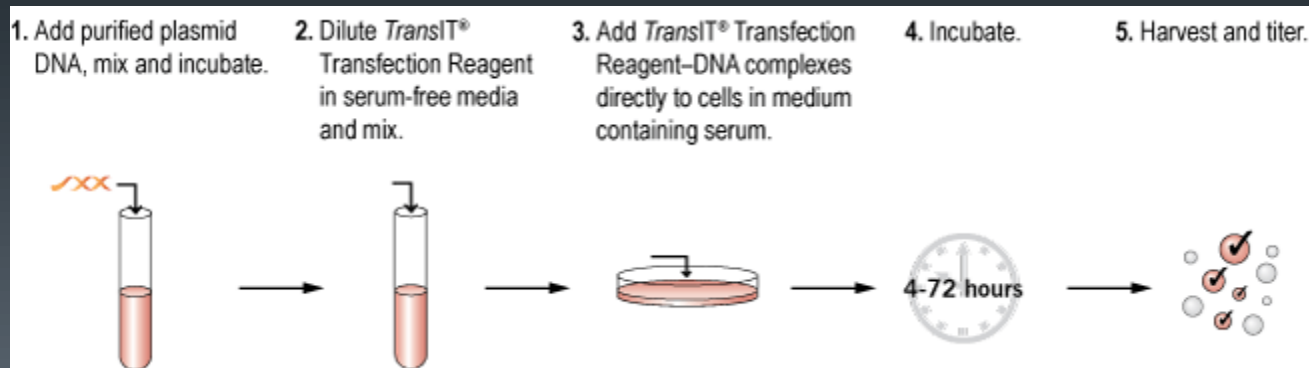
# 1<sup>st</sup> Step - Cell Expansion

- We first need a master cell bank
- Cell Expansion in adherent cell culture
- Adherent cell factories
- Plate HEK-293 cells
- Plasmid DNA should be of high quality and free of RNA contaminants.



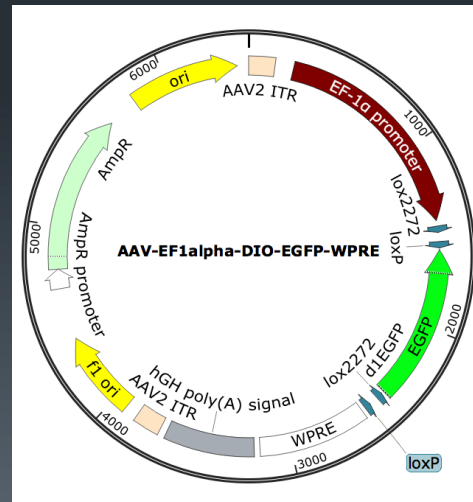
## 2<sup>nd</sup> Step – Pre-Plasmid Transfection

- Transfection with 2-4 plasmid vectors
  - pFdelta6, pRV1, pH21, and AAV
- Optimized harvest and transfection



# 3<sup>rd</sup> step – Post-Plasmid Transfection

- Virus production phase post-transfection
- Filter the transfection mixture into a 50 ml tube.
- Optimized harvest and transfection



## 4<sup>th</sup> Step – Harvest AAV

- 72 hours after transfection, remove media from cell culture plates and discard.
  - All waste should be treated with Virkon solution or other suitable disinfectant.
- Virus extraction, clarification, and concentration
- Recovery and stabilization of virus from cell-culture system
- Incubate at 37°C for 1 hour



# 5<sup>th</sup> Step- Purify & Concentrate AAV's

- Setup HiTrap heparin columns using a peristaltic pump so that solutions flow through the column. It is important to ensure no air bubbles are introduced into the heparin column.

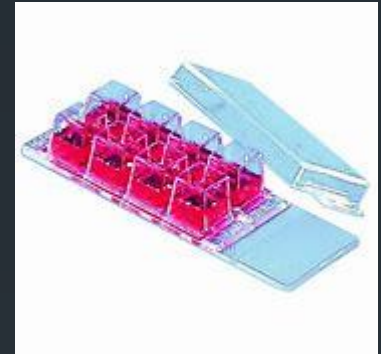


- Purification Process ahead of DS Formation
- Concentrate vector using Amicon ultra-4 centrifugal filter units with a 100,000 molecular weight cutoff.



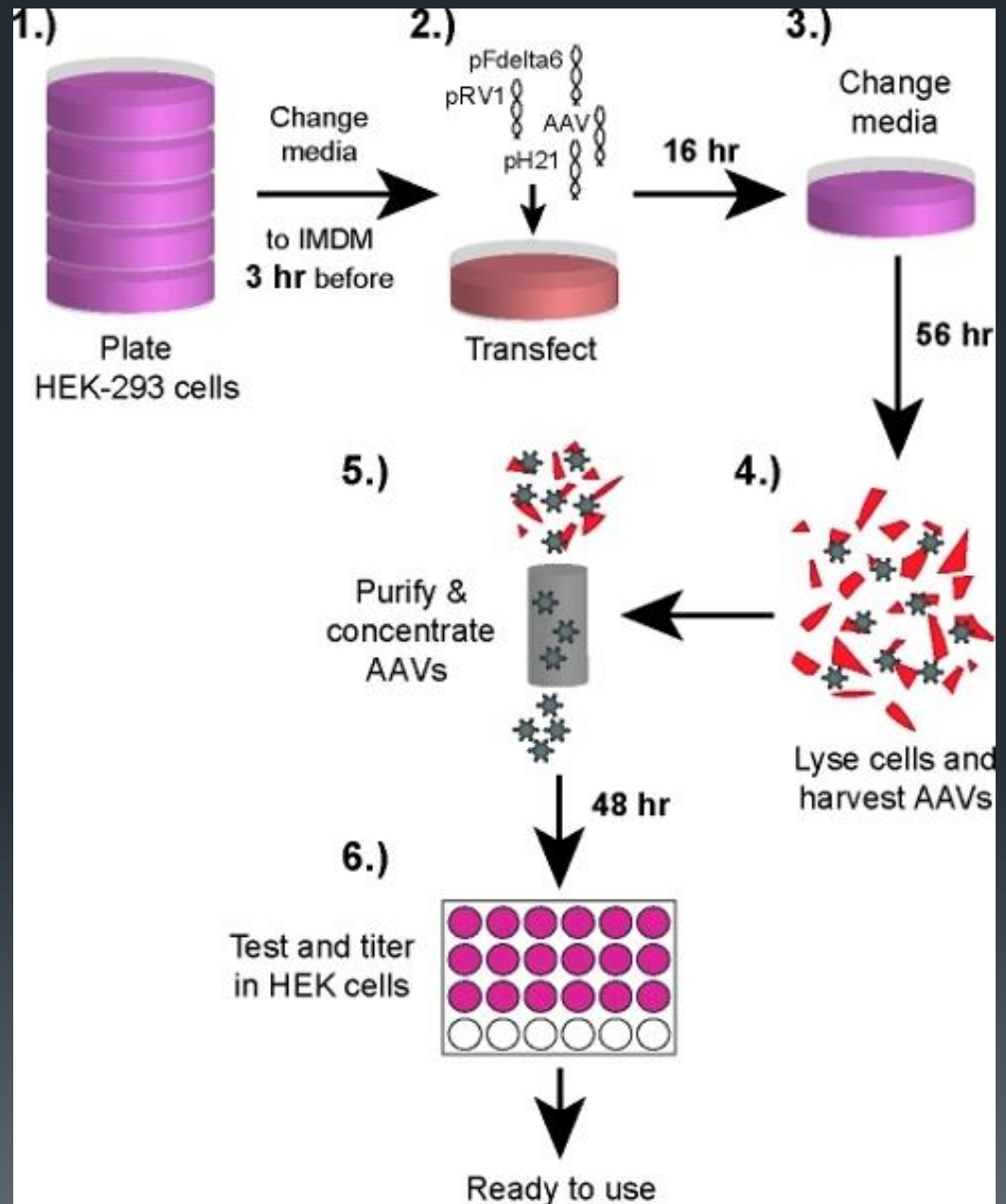
## 6<sup>th</sup> step – Test and titer in HEK cells

- Vector should be aliquoted and stored at negative 80°C until required.
- This requires a promoter that is active in Hek293 cells driving a reporter gene or an immunocytochemically detectable gene product.

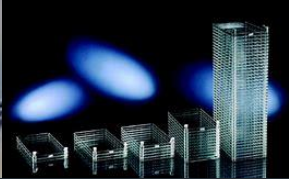


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# Summary -







AAV



1. Add purified plasmid DNA, mix and incubate.

2. Dilute *TransIT*<sup>®</sup> Transfection Reagent in serum-free media and mix.

3. Add *TransIT*<sup>®</sup> Transfection Reagent-DNA complexes directly to cells in medium containing serum.

4. iter.

